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(54) **Viral replicons and viruses dependent on inducing agents**

(57) The present invention relates to inducible viral replicons as regulatable live-attenuated vaccine candidates. In particular, it relates to the HIV-rTA replicon whereby, aiming for regulated expression of the replicon, the rTA gene has been inserted into the HIV1-Nef gene (i.e. under control of Nef regulatory elements), the TetO elements have been inserted into one or either LTRs, and the NF-Kappa-B sites of said LTRs have of have not been mutated. Aiming for further attenuation,

the TAR hairpin and eventually also the Tat gene have been mutated. A conditional cytotoxic gene (e.g. HSV-tk) can eventually also be inserted into the replicon to provide for improved biosafety. Said replicon is intended to be used as live-attenuated HIV1 vaccine, but forced evolution (i.e. genetic drift of HIV due to the relatively poor fidelity of HIV1 RT) could be used to improve the present mutants, as well as to select for modified/improved rTA-TetO regulatory systems.

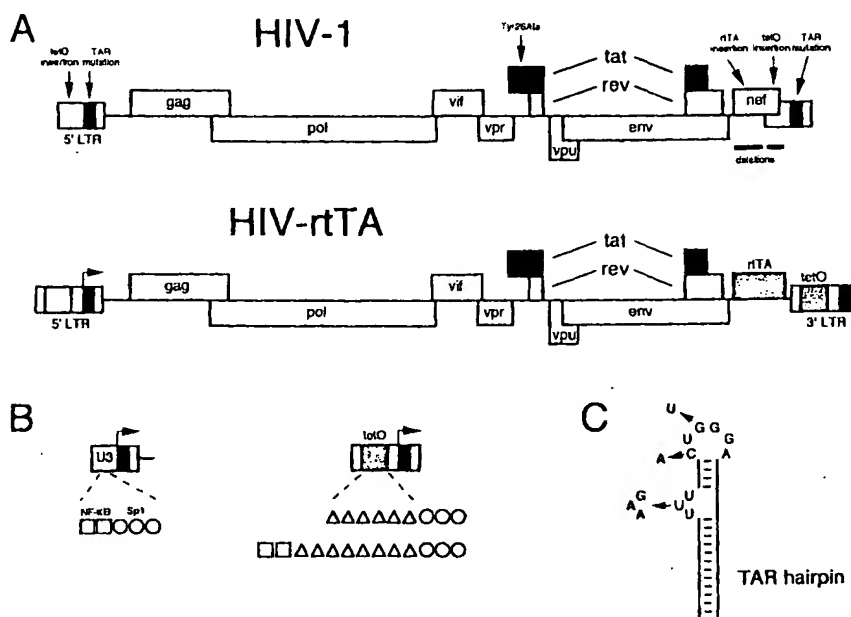


Fig. 1

Description

[0001] The present invention relates to the field of molecular biology of pathogens, in particular viruses and more in particular human immunodeficiency virus. It relates to methods for producing replicons and/or viruses dependent on inducing agents, said viruses and/or replicons as well as uses of such replicons and/or viruses in the production of vaccines, in particular live-attenuated vaccines. Live-attenuated virus vaccines (such as vaccinia, polio and measles) have been enormously successful and have made a dramatic and historic impact on public health. However, for the human immunodeficiency virus type 1 (HIV-1) safety concerns remain about either the reversion of attenuated vaccine strains to virulent phenotypes or the induction of fulminant infection in (immunocompromised) individuals. Testifying to the genetic instability of such strains is the recent demonstration that the HIV-1 delta3 vaccine candidate, which contains 3 deletions in non-essential parts of the genome, is able to regain full replication capacity within four months of replication in tissue culture (Berkhout et al., 1999). In addition, it has been reported recently that replication of deletion variants of the simian immunodeficiency virus (SIV) increased after several years in some infected monkeys, concomitant with the onset of AIDS (Baba et al., 1999). Furthermore, although there is some evidence that attenuated HIV-1 variants lacking the *nef* gene result in a benign course of infection in humans (Deacon et al., 1995), a decline in CD4+ T-cell numbers has been reported recently for some of these individuals, which is an early sign that these persons could develop AIDS (Dyer et al., 1999; Greenough et al., 1999). These results have forced the development of subunit or inactivated virus vaccines, but these vaccines have not elicited the potent broad-based immune responses or long-term memory necessary to confer life-long protection in immunized individuals [reviewed in (Paul, 1995)]. It is for this reason that live-attenuated HIV vaccine approaches are still being considered. Replicating virus vaccines demonstrated superior performance in AIDS vaccine trials. It has been repeatedly demonstrated that macaques or chimpanzees persistently infected with genetically attenuated, non-pathogenic isolates of SIV or HIV-1, respectively, strongly resist a subsequent challenge with pathogenic virus (Shibata et al., 1997; Wyand et al., 1996; van Rompay et al., 1995; Almond et al., 1995; Daniel et al., 1992; Lohman et al., 1994; Stahl-Hennig et al., 1996; Johnson et al., 1999). However, to satisfy safety concerns, the ideal vaccine strain should replicate only to the extent that is needed for immunogenicity. Towards the construction of the next generation of safe, genetically stable HIV-1 variants as live-attenuated AIDS vaccine, we now report the construction of a HIV-1 variant of which the replication depends on the addition of an inducing agent such as the non-toxic, selective effector doxycycline (dox). Thus the invention provides an inducible vi-

ral replicon, comprising all viral sequences which are essential in cis for replication under direct or indirect control of at least one inducible repressor and/or activator. The invention is exemplified by the preferred embodiments relating to Human Immunodeficiency Virus (HIV). However, the invention will be applicable to other pathogens, in particular viral pathogens, of which it is important that they replicate in order to obtain an efficacious immune response, but for which it is also important that said replication does not go beyond the level required for said immune response. A replicon is defined as a nucleic acid molecule capable of replication in a suitable environment, such as a permissive cell, because it has all the necessary elements for replication in such an environment. We call it a replicon, because it will not always be directly derived from the nucleotide sequences of the original pathogen, for instance in the case of single stranded DNA viruses, RNA viruses, etc. Typically, in order to manipulate nucleic acids, double stranded forms are necessary, typically double stranded DNA forms. Therefore preferred replicons will be double stranded DNA nucleic acids in at least one stage of their life cycle.

[0002] A replicon is also intended to reflect that the actual pathogen, or its attenuated live vaccine relative, usually comprises more than just nucleic acid. The nucleic acid is typically packaged into a (viral) particle. Therefore the replicon preferably also encodes a functional packaging signal, allowing for the nucleic acid in its wild-type-like form (RNA in the case of a retrovirus, etc.) to be packed into a viral particle. In order for the replicon to be able to replicate in a host, it is preferred that said replicon also carries the structural genes for the proteins of the envelope and/or capsid, be it in wild-type format or in a somewhat different format (reduced or enhanced target binding, etc.).

[0003] In order to be able to regulate the amount of replication necessary for eliciting a good immune response without any replication beyond that level, according to the invention at least one gene essential for said replication is placed under the control of an inducible repressor/activator. In order to prevent leakage, it is preferred to have a combination of essential genes under such control and it is even more preferred to have at least two different repressor/activator combinations in control of at least one, but preferably more than one gene essential for replication. In most (viral) pathogens a number of genes is essential for replication, but most of them also have a sort of "master switch", usually an early gene, usually transactivating other genes. A first candidate to put under direct control of a repressor/activator is of course such a master switch, which then indirectly provides control over the other essential genes for replication. Still it is even then preferred to put at least one other essential gene under control of an inducible repressor/activator. Such a master switch is not required for 'simple' viral genomes such as HIV-1 that are under control of a single transcription unit.

[0004] As stated before the replicon is preferably a viral replicon which is derived from a human immunodeficiency virus. Typically such a replicon would be an infectious double stranded DNA clone of an HIV strain. Preferably said HIV strain is already an attenuated strain, or is made into an attenuated strain by introducing mutations, such as functional deletions, e.g. those described herein. Any repressor/activator elements that are inducible are in principle applicable in the present invention. Typically when they are used as a single element they should not have leakage (meaning low base levels of gene expression) in the repressed or unactivated state. In the case of double or more inducible controls, such leakage becomes less important, although essentially no leakage is still highly preferred. A good system for inducible control is the combination of the Tet-operon, together with doxycyclin as the inducing agent. Thus the invention also provides a viral replicon wherein said inducible repressor and/or activator comprises a Tet operon or a functional equivalent thereof. This operon and its necessary elements are as such known and further described herein below. A functional equivalent thereof is an element that is capable of repression and/or activation in essentially the same manner as the Tet operon. Typically this would be highly homologous variations of said operon. As a safety valve, it would be advantageous to provide the replicon with a suicide gene that can be activated when unwanted effects occur such as replication beyond what is necessary for an immune response or rescue by wild type virus, etc. Such a suicide gene is e.g. HSV-tk, which can be induced by adding gancyclovir or a functional equivalent thereof. Upon induction said gene will kill the infected cell, and thereby inhibit further replication and infection of other cells. Thus in yet another preferred embodiment the invention provides a replicon according to the invention which further comprises a suicide gene.

[0005] As stated herein before, the replicon is preferably under control of at least a Tet operon, which allows for replication in the presence of doxycyclin. Thus the invention also provides a replicon according to the invention which can be induced to replicate by the presence of doxycyclin or a functional analog thereof.

In the present context a functional analog of doxycyclin is a molecule capable of removing repression or initiating activation of the genes under control of the activator and/or repressor present in the replicon.

In order to attenuate the HIV replicon and/or the resulting virus it is preferred that the replicon is provided with a functional deletion of the TAR-element. Thus in yet another preferred embodiment the invention provides a replicon according to the invention, which further comprises an inactivated TAR element.

[0006] In order to attenuate the HIV replicon according to the invention it is preferred to functionally delete the Tat element. Thus the invention also provides a replicon according to the invention, which further comprises an inactivated Tat element. Preferably both elements

mentioned above are functionally deleted. Functional deletion means that at least their function in the replication of the replicon is at least partially inhibited. Essential genes for replication typically should not be completely dysfunctional.

Proteins necessary for removing repression or initiating activation elements which are present upstream of the essential genes to be put under control should be encoded by the replicon and should be inserted preferably in a non-essential gene. Thus the invention also provides in a preferred embodiment a replicon according to the invention wherein at least one functional part, preferably an rTA gene, of said inducible repressor and/or activator is inserted into the *nef* gene. The functional part in this case of course refers to any proteinaceous substance capable of activating or derepressing the element in control of the essential gene. Preferably space is created for the sequence encoding said proteinaceous substance. Thus the invention also provides a replicon in which at least part of the *nef* gene is deleted to create space for insertion.

To further attenuate a replicon according to the invention further elements of the wild-type virus can be functionally deleted. Thus the invention further provides a replicon according to the invention, in which at least one NF-kB element has been deleted. It is preferred that the motif to be activated is a tetO motif, preferably present in an LTR. Thus the invention also provides a replicon, which comprises at least one tetO motif in at least one functional LTR.

It is preferred to have more than one element before an essential gene. Thus the invention also provides a replicon which comprises at least 2, 4, 6, or 8 such elements in at least one functional LTR.

The LTR is preferably modified to avoid reversion to wild type virus.

[0007] The invention further provides methods using the replicons to produce dependent viruses, meaning viruses needing an inducing agent in order to be able to replicate. Thus the invention provides a method for producing a virus dependent on an inducing agent for replication, comprising providing a permissive cell with a replicon according to the invention, culturing said cell in the presence of said inducing agent and harvesting said dependent virus from said culture. Again such methods are preferably applied to HIV. Thus the invention provides a method in which said dependent virus is a human immunodeficiency virus, preferably an attenuated virus.

[0008] The preferred inducing agent is again doxycyclin. Thus yet another preferred embodiment is a method in which said inducing agent is doxycyclin or a functional analog thereof. Also part of the present invention are viruses produced by said methods or which can be produced by said methods. Thus the invention also provides a virus dependent on an inducing agent for replication obtainable by a method according to the invention, preferably again a human immunodeficiency virus,

again preferably attenuated.

[0009] The viruses will find an important application in vaccination. The invention thus also provides a vaccine comprising a replicon according to the invention and/or a virus according to the invention, an amount of inducing agent and optionally a suitable adjuvant.

[0010] Of course the vaccine may comprise a single dosage unit, but it may also comprise the inducing agent separately, or it may be made on the spot from a replicon and/or virus that are reconstituted with a liquid excipient such as saline, optionally together with an adjuvant and/or an inducing agent. Viral vaccines are well known in the field. General rules of thumb applicable to known vaccines will also apply to the vaccines of the present invention. Doses will be found through the normal dose finding studies performed during (pre)clinical trials, e.g. by simple titration of the amount of doxycycline as inducing agent. The vaccine may be sufficient on its own, but it may also be used in addition to other vaccines, etc. The inducing agent may be needed over a longer period of time and can then be provided separately. Again the preferred vaccine is one for prophylaxis of infection with a human immunodeficiency virus.

The invention also provides the use of said vaccine in that it provides a method for the prophylaxis of AIDS, comprising administering a vaccine according to the invention to a patient and allowing for viral replication for a limited time by providing said inducing agent. Booster vaccinations are possible by simple readmission of the said inducing agent at later times.

The invention also provides a method for the controlled replication of a virus or a viral replicon comprising providing a permissive cell with a replicon or a virus according to the invention, culturing said cell in the presence of said inducing agent and manipulating the amount of inducing agent present.

Detailed description.

[0011] As stated before the replication of a viral replicon was put under control of a repressor and/or activator system. In the examples this was done by incorporation of the Tet-system into the HIV-1 genome.

[0012] Several eukaryotic systems for inducible gene expression have been reported, but the Tet-induced regulatory circuit has some unique properties for incorporation into HIV-1 (Gossen et al., 1993; Gossen & Bujard, 1992; Baron et al., 1999). This Tet-system has found wide application and strict and graded regulation of gene expression has been reported in many experimental set-ups, for example, in the breeding of transgenic animals and in gene therapy approaches. Another advantage of these well-characterized regulatory elements from an evolutionary distant organism such as *E. coli* is that one can establish truly monospecific regulatory circuits in higher eukaryotic cells, thereby limiting the danger of unwanted side effects. This system is based on two elements from the *E. coli* tet operon, the

tetracycline-inducible repressor protein (TetR) that has been converted into a eukaryotic transcriptional activator (tTA or rTA), and the tetO operator DNA sequence. We designed a novel strategy to impose regulation on HIV-1 gene expression and replication with the Tet-system, such that an exogenous agent (dox) can be used to reversibly turn on and off viral replication.

Construction of HIV-rTA viruses. The full-length, infectious HIV-1 molecular clone pLAI was used for construction of an HIV-rTA virus genome in which the TAR-Tat axis (red in Figure 1A) was replaced by the TetO-rTA elements (green). In general, we took a conservative approach with regard to the type of mutations that were introduced in the HIV-1 genome in order to minimize the chance to inactivate unknown replicative signals.

[0013] **TAR and Tat inactivation.** First, the TAR element was inactivated by mutation of nucleotides in the single-stranded bulge and loop domains (Figure 1C). Combination of the bulge and loop mutations produces a fully inactive TAR motif because even point mutations in one of these single-stranded TAR domains have a dramatic effect on TAR-function in Tat-mediated LTR transcription and virus replication (Berkhout & Jeang, 1991; Berkhout & Jeang, 1989; Berkhout & Klaver, 1993). We did not introduce more gross sequence changes or even deletions in TAR because this sequence is also essential for virus replication as repeat-R region during strand transfer of reverse transcription (Berkhout et al., 1995). Although we demonstrated previously that the TAR element of the 5'LTR is inherited in both LTRs of the viral progeny, the inactive TAR motif was inserted in both LTRs to minimize the chance of a reversion to the wild-type virus by a recombination event (Klaver & Berkhout, 1994).

[0014] Inactivation of the Tat protein was accomplished by introduction of the Tyr26Ala pointmutation. This single amino acid change results in a complete loss of Tat transcriptional activity and viral replication capacity (Verhoef et al., 1997). The corresponding codon change (UAU to GCC) was designed to restrict the likelihood of simple reversion to the wild-type amino acid, which requires at least two substitutions (Verhoef & Berkhout, 1999). It has been suggested that Tat may play additional roles in the replication cycle besides its transcriptional function (Huang et al., 1994; Harrich et al., 1997; Ulich et al., 1999). Thus, Tat may facilitate HIV-rTA replication even in the absence of an intact TAR element, and we therefore also made viruses with the wild-type tat gene. These constructs will be referred to as Y (tyrosine mutant) and W (wild-type).

[0015] **rTA and tetO insertion.** Two deletions were introduced in the *nef* gene create space for the insertion of the non-viral elements (Figure 1A). A 250-nt upstream fragment was removed, and a 200nt fragment overlapping the U3 region of the 3'LTR. This U3-deletion will be inherited by the viral progeny in both LTRs. The exact borders of the U3 and Nef deletions were carefully cho-

sen such that important *cis*-acting sequences for virus replication were not removed. In particular, we maintained approximately 80-nt around the 5' end of the 3'LTR (Figure 1A). This region encodes multiple sequence elements that are critical for reverse transcription (Ilyinskii & Desrosiers, 1998) and integration (Brown, 1997). In fact, we tried to mimic spontaneous deletions that have been observed in the *nef*/U3 region of several HIV and SIV variants in a variety of replication studies, including *in vivo* experiments (Kirchhoff et al., 1994; Fisher & Goff, 1998; Ilyinskii et al., 1994; Kirchhoff et al., 1995). As preparation for the insertion of the exogenous rTA gene into the position of the *nef* gene, a short synthetic sequence was inserted that provides a translational start codon in an optimal sequence context (CCAUGU, (Kozak, 1989) and convenient restriction enzyme recognition sites. The rTA gene was inserted as *XcmI*-*XbaI* fragment in this polylinker segment, in frame with the optimized start codon. The splice acceptor that is located just upstream of the *nef* gene was maintained, and rTA translation should occur from the subgenomic mRNA that was originally meant for expression of the Nef protein.

[0016] To identify the optimal configuration of an LTR promoter with rTA-responsive tetO elements, we first performed transient transfection studies with a variety of LTR-luciferase constructs (Verhoef *et al.*, manuscript in preparation). We varied the number of tetO motifs (2, 4, 6, or 8) that were inserted upstream of the three Sp1 binding sites of the HIV-1 LTR promoter. We also tested constructs with and without the two upstream NF- κ B elements. The two promoters that provided most robust dox-induced transcription were selected for insertion into the HIV-1 genome and these LTRs are schematically depicted in Figure 1B. They will be referred to as **K** (NF- κ B + 8 tetO + Sp1) and **S** (6 tetO + Sp1). Although insertion into the U3 region of the 3'LTR will be sufficient to produce a mutant progeny, we also introduced the tetO motifs in the 5'LTR to generate molecular clones of which the initial round of gene expression in transfected cells is also regulated in a dox-dependent manner. Thus, both LTRs were modified, and this was done in the wild-type and mutant Tat background, resulting in four HIV-rTA constructs: **KWK**, **KYK**, **SWS**, and **SYS**. All HIV-rTA molecular clones have the TAR inactivation and rTA insertion in common, but they differ in the status of the *tat* gene and the type of tetO insert. Of these virus variants, **KWK** is most wild-type-like because it maintained the NF- κ B sites and a wild-type Tat protein, and **SYS** is the most minimal HIV-rTA version.

HIV-rTA replicates in a doxycycline-dependent manner.

[0017] The four pLAI plasmids were individually transfected into the SupT1 T cell line to test for their replication capacity. The culture was maintained at varying dox levels, and virus replication was monitored by measur-

ing the amount of CA-p24 produced in the culture medium (Figure 2). In the presence of optimal dox levels (1000 ng/ml), we measured profound replication of all four HIV-rTA viruses. No virus replication was observed in the absence of dox, indicating that replication is strictly dependent on the inserted Tet-system. The Tet-system is ideally suited to modulate the level of transcriptional activation in a step-wise manner by reducing the amount of dox (Baron et al., 1997). Indeed, replication of the HIV-rTA viruses can also be modulated at sub-optimal concentrations of the inducing dox reagent (Figure 2). A progressive reduction in replication rates of all four rTA-viruses was observed at 300 and 100 ng/ml dox, and virus replication was nearly abolished at 30 ng/ml. These combined results demonstrate that the HIV-rTA viruses replicate in a strictly dox-dependent manner and that the rate of replication can be fine-tuned by simple variation of the dox-concentration.

[0018] The transfected SupT1 cells were killed within 1 week by massive virus-induced syncytia, and CA-p24 production levels reached values that are similar to what is observed in regular infections with the wild-type LAI virus.

Nevertheless, the HIV-rTA variants have a significantly reduced fitness because they showed delayed replication in transfections with less DNA (results not shown). Although the four viruses appear to have a similar replication capacity, this can be measured more appropriately in subsequent infection studies. Indeed, we managed to passage all four HIV-rTA viruses as cell-free inoculum onto fresh, uninfected T cells, and a spreading infection was sustained for at least 5 weeks (5 passages). From these infection experiments the following ranking order of replication was apparent: **KWK** > **KYK**, **SWS** > **SYS**.

[0019] HIV-rTA vaccine viruses should be able to replicate in primary cells. The LAI molecular clone used in these studies represents a primary isolate that is able to efficiently infect primary cells (Wain-Hobson et al., 1991; Peden et al., 1991), but a complication of our design is that we removed the *nef* gene, which contributes to virus replication in primary cell types (de Ronde et al., 1992). We transfected pooled peripheral blood mononuclear cells (PBMC) by means of electroporation with 20 μ g of the molecular clones and measured CA-p24 production in the culture supernatant for up to two weeks (Figure 3). All four HIV-rTA variants replicated in the presence of 1000 ng/ml dox, whereas no replication was detectable without dox. The ranking order of replication in PBMCs (**KWK** > **KYK** > **SWS** > **SYS**) is very similar to that observed in SupT1 cells.

Turning virus replication on and off in a reversible manner.

[0020] Subsequent tests were performed with the **SWS** virus in SupT1 infections (Figure 4). First, we repeated the dox-response experiment. In this more sen-

sitive infection experiment, it is obvious that the sub-optimal amount of 100 ng/ml dox allows only a low level of replication that is not sufficient to support a spreading infection (Figure 4A). We next analyzed virus replication kinetics when dox was added 3 days after infection of the cells (Figure 4B). This resulted in a delay of virus production of approximately 3 days. In the absence of dox, the HIV-rtTA virus can still infect cells, reverse transcribe its RNA genome and integrate the DNA into the host genome. In other words, the provirus form can be established. This latently infected cell will remain in the culture, and can be activated by dox after three days. An additional feature of the Tet-system is that it provides reversible regulation, and this was tested in the replication assay (Figure 4C). SupT1 cells were infected with the SWS virus and cultured in the presence of dox. At day 3, the cells were washed to remove extracellular dox, and resuspended in medium either with or without dox. Indeed, replication can be stopped abruptly by removal of dox. These combined results confirm that replication of the HIV-rtTA virus is absolutely dependent on dox, and the level of virus replication can be strictly controlled in a graded and reversible manner.

Safety issues.

[0021] Several assays were performed to analyze different safety aspects of the HIV-rtTA variants. First, we screened for leaky virus replication in the absence of dox. For instance, the cell cultures that were transfected with the four different HIV-rtTA constructs (Figure 2) were maintained without dox for a prolonged period of time, but no virus production was measured in these four cultures up to day 52, at which point we stopped the experiment. Similarly, no replicating virus was observed in primary cells without dox (Figure 3). In addition, SupT1 cultures in which virus spread was ongoing in the presence of dox were 'turned off' by removal of dox (see e.g. Figure 4C for the SWS virus), without any sign of virus production. These experiments may be viewed as the first safety tests of these vaccine strains.

[0022] As an additional safety test, we analyzed the sensitivity of the HIV-rtTA virus to antiretroviral drugs that are in current clinical use. Because we did not alter the basic set of viral genes in HIV-rtTA, including the genes encoding Protease (Pro) and Reverse Transcriptase (RT), these viruses are expected to remain fully sensitive to well-known drugs that target these essential enzymes. As shown in Figure 4D, replication of the dox-dependent SWS virus can be inhibited efficiently either by 3'-azido, 3'-deoxythymidine (AZT, a nucleoside RT-inhibitor) or Saquinavir (SQV, a Pro-inhibitor).

Discussion.

[0023] We have incorporated the Tet-transcriptional system in the HIV-1 genome such that virus replication can now be controlled from the outside by addition of a

non-toxic inducer molecule such as doxycycline (dox). Specifically, we constructed replicating HIV-1 variants with inactivating mutations in both arms of the Tat-TAR axis through replacement with the rtTA-tetO elements of the Tet-system. Replication experiments in a T cell line and primary cells convincingly demonstrate that we have successfully designed dox-dependent HIV-1 variants. Replication of these designer HIV-rtTA viruses is regulatable in a graded and reversible manner. Although 'leakiness' has been a problem in some protocols using the rtTA system, we have not observed any virus replication in the absence of dox. One possible explanation is that expression of the rtTA *trans*-activator in the HIV-rtTA system is fully dependent on the presence of dox. Thus, an autoregulatory loop may have been established that resembles the natural TAR-Tat axis. This mechanism may restrict leakiness or dox-independent replication, thereby providing a significant additional safety feature.

[0024] The HIV-rtTA viruses have some unique properties make them ideal reagents for a variety of biological experiments. The most obvious application for such a virus is in the field of live-attenuated vaccines, and a similar approach may be used to put control over other retroviral pathogens (e.g. HIV-2, HTLV-I), pararetroviruses (e.g. HBV), or DNA viruses (e.g. Herpesvirus or Adenovirus). The HIV-rtTA viruses improve the current generation of live-attenuated HIV-1 variants as potential vaccine strains because the conditional replication adds a unique safety feature. The SYS variant has the most minimal 'genotype': TAR⁻, Tat⁻, delta-U3, delta-NF-kB, delta-*nef*, but it should be possible to delete in addition some of the 'accessory' genes such as *vpr*, *vpu* and/or *vif*. HIV-rtTA vaccine viruses should be able to induce a protective immune response, after which replication can be turned off, such that the virus will be stably non-pathogenic. The HIV-rtTA viruses can still be inhibited by antiviral drugs that are in clinical use, and this was demonstrated for the RT-inhibitor AZT and the Pro-inhibitor Saquinavir. These viruses await extensive replication tests to verify their genetic stability, followed by animal tests to screen for their pathogenic potential and their ability to induce a protective immune response.

[0025] Because the TAR RNA and *tat* gene may have become non-essential parts of the HIV-rtTA genome, these elements may now be 'free' to evolve. If these elements have indeed no other function in the viral replication cycle, one would predict that they would eventually be lost by the accumulation of mutations and/or deletion. This further reduces that likelihood of a wild-type-like reversion, thereby making the vaccine strain more safe. However, the situation may be more complex as additional roles have been proposed for both motifs. This is most obvious for the TAR motif, which is part of the R (repeat) region that is critical in strand transfer during reverse transcription. But TAR has also been reported to contribute to RNA packaging in virion particles [reviewed in (Berkhout, 1999)]. The Tat protein has also

been implicated in non-transcriptional roles, e.g. during mRNA translation and the process of reverse transcription (SenGupta et al., 1990; Huang, Joshi, Willey, Orenstein, and Jeang, 1994; Harrich, Ulich, Garcia-Martinez, and Gaynor, 1997; Cullen, 1986). Prolonged culture experiments and the analysis of revertant viruses will provide more insight into some of these possibilities.

[0026] The HIV-1 TAR-Tat axis was successfully replaced by the tetO-rTA system, and the latter elements have become essential viral functions. This also adds an important safety feature because it will preclude the spontaneous loss of the new viral elements by deletion, an event that occurs frequently with exogenous sequences that are inserted in a (retro)viral genome. Thus, this feature further enhances the genetic stability of vaccine strains based on HIV-rTA. On the other hand the current HIV-rTA variants do not yet replicate optimally, and this is particularly true for the most minimal SYS variant that lacks a functional Tat gene and NF-kB sites. It is therefore probable that continued replication will lead to improvement of this new HIV-1 transcriptional axis by selection of spontaneous up-mutants. The beauty of working with HIV is that even if a poorly replicating virus is identified, the error-prone nature of the RT enzyme will allow for the generation of faster-replicating variants by a method termed forced evolution (Klaver & Berkhout, 1994; Berkhout & Das, 1999). This evolutionary refinement of the initial designer HIV-rTA variants provides a powerful method to select for fast-replicating, dox-dependent HIV-1 variants. This evolutionary approach will identify modified forms of the rTA protein or the tetO sites that are better suited for their new role in virus replication.

[0027] We expect that we can even use the enormous evolutionary capacity of HIV-1 to select for rTA elements with altered substrate-specificity by gradually changing dox for other dox-like derivatives in the culture medium. Thus, the virus will help us to find better tetO-rTA reagents that can subsequently be useful in biological settings that require specific regulation of gene expression (e.g. transgenic mice, gene therapy). We plan to rigorously test the possibility to perform genetics with the 'prokaryotic' Tet-system in this eukaryotic (viral) background.

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Legends to the figures

[0029] Figure 1. Design of a Tetracycline-dependent HIV-1. Panel A shows the HIV-1 genome and multiple modifications that were introduced to construct HIV-rtTA. Details of the mutations are provided in the text, see also panels B and C for the LTR modifications. In brief, we inactivated the TAR-Tat transcriptional axis (marked in red) and replaced it by the Tetracycline-inducible tetO-rtTA system (marked in green). Inactivation of the TAR and Tat is marked by crosses through the motifs. The genome maps are not drawn to scale, but the genome size of HIV-rtTA is larger than that of HIV-1. The RNA genome of HIV-1 LAI is 9229-nt, and HIV-rtTA is either 9767-nt (the S.S variants) or 9875-nt (K.K variants). Panel B provides some details of the tetO insertions in the LTR promoter. The U3 region of the wild-type LTR (left) encodes 2 NF-kB sites (squares) and 3 Sp1 sites (circles). The modified LTR (right) contains either 6 or 8 tetO operators (green triangles) upstream of the Sp1 sites. The 6 tetO variant only has the Sp1 sites in mutant S, whereas both NF-kB sites are present upstream of the 8 tetO operators in mutant K. The arrow marks the transcription start site at the U3-R border, which also is the start site of the TAR hairpin. Panel C shows the TAR hairpin structure and the inactivating mutations that were introduced in the bulge (triple-nucleotide substitution) and in the loop (two point mutations). These mutations should disrupt binding of the viral Tat protein and the cellular cyclin T co-factor, respectively (Dingwall et al., 1989; Wei et al., 1998).

[0030] Figure 2. Doxycycline-controlled replication of the HIV-rtTA viruses. The SupT1 T cell line was electroporated with 10 µg of the indicated molecular clones, and cells were cultured in without or with an increasing concentration of dox (0 to 1000 ng/ml range). Virus production was measured by CA-p24 elisa on culture supernatant samples.

[0031] Figure 3. Doxycycline-dependent replication of HIV-rtTA viruses in primary cells. PBMCs were electroporated with the four individual HIV-rtTA constructs (20 µg), and the cultures were maintained without or with (1000 ng/ml) dox. Fresh uninfected cells were added immediately after transfection and at day 6 post infection. Virus production was measured by CA-p24 elisa on culture supernatant samples.

[0032] Figure 4. Replication of HIV-rtTA can be turned on and turned off. These experiments were performed with the SWS virus, but similar results have been obtained with the other HIV-rtTA variants. We used the SWS virus (2200 ng CA-p24) to infect 6 x 10⁶ SupT1 cells at day 0. Panel A shows the replication potential with 0, 100 and 1000 ng/ml dox. In panel B, we analyzed the effect of delayed addition of dox (1000 ng/ml) at day 3 after infection. In panel C, the infected cells were grown in 1000 ng/ml dox for 3 days, at which point the cells were washed and incubated in the absence or presence of dox. In panel D, infected cells were maintained in the presence dox and we tested the effect of the Protease-inhibitor Saquinavir (200 nM) and the RT-inhibitor AZT (1 µg).

Claims

1. An inducible viral replicon, comprising all viral sequences which are essential in cis for replication under direct or indirect control of at least one inducible repressor and/or activator.
2. A viral replicon according to claim 1 which is derived from a human immunodeficiency virus.
3. A viral replicon according to claim 2 which is derived from an infectious human immunodeficiency virus clone.
4. A viral replicon according to any one of claims 1-3, wherein said inducible repressor and/or activator comprises a Tet operon or a functional equivalent thereof.
5. A replicon according to any one of claims 1-4 which encodes an attenuated virus.
6. A replicon according to any one of claims 1-5 which further comprises a suicide gene.
7. A replicon according to any one of claims 1-6, which can be induced to replicate by the presence of doxycyclin or a functional analog thereof.
8. A replicon according to any one of claims 2-7, which further comprises an inactivated TAR element.
9. A replicon according to any one of claims 2-8, which further comprises an inactivated Tat element.
10. A replicon according to any one of claims 2-7, wherein at least one functional part, preferably an rtTA gene, of said inducible repressor and/or activator is inserted into the nef gene.
11. A replicon according to claim 10 in which at least

- part of the nef gene is deleted to create space for insertion.
12. A replicon according to any one of claims 2-11, in which at least one NF-kB element has been deleted. 5
 13. A replicon according to any one of claims 2-12, which comprises at least one tetO motif in at least one functional LTR. 10
 14. A replicon according to claim 13, which comprises at least 2, 4, 6, or 8 such elements in at least one functional LTR. 15
 15. A replicon according to any one of claims 2-14, wherein at least one LTR is modified to avoid reversion to wild type virus.
 16. A method for producing a virus dependent on an inducing agent for replication, comprising providing a permissive cell with a replicon according to any one of claims 1-15, culturing said cell in the presence of said inducing agent and harvesting said dependent virus from said culture. 20 25
 17. A method according to claim 16, in which said dependent virus is a human immunodeficiency virus.
 18. A method according to claim 16 or 17 in which said virus is an attenuated virus. 30
 19. A method according to any one of claims 16-18, in which said inducing agent is doxycyclin or a functional analog thereof. 35
 20. A virus dependent on an inducing agent for replication obtainable by a method according to any one of claims 16-19. 40
 21. A virus according to claim 20, which is a human immunodeficiency virus.
 22. A virus according to claim 20 or 21 which is an attenuated virus. 45
 23. A vaccine comprising a replicon according to any one of claims 1-15 and/or a virus according to any one of claims 20-22, an amount of inducing agent and a suitable adjuvant. 50
 24. A vaccine according to claim 23 whereby said inducing agent is provided separately.
 25. A vaccine according to any one of claims 23-24, wherein said replicon and/or said virus is derived from a human immunodeficiency virus. 55
 26. A method for the controlled replication of a virus or a viral replicon comprising providing a permissive cell with a replicon according to any one of claims 1-15, or a virus according to claim 20, 21 or 22, culturing said cell in the presence of said inducing agent and manipulating the amount of inducing agent present.
 27. A method for the prophylaxis of AIDS, comprising administering a vaccine according to any one of claims 23-25 to a patient and allowing for viral replication for a limited time by providing said inducing agent.

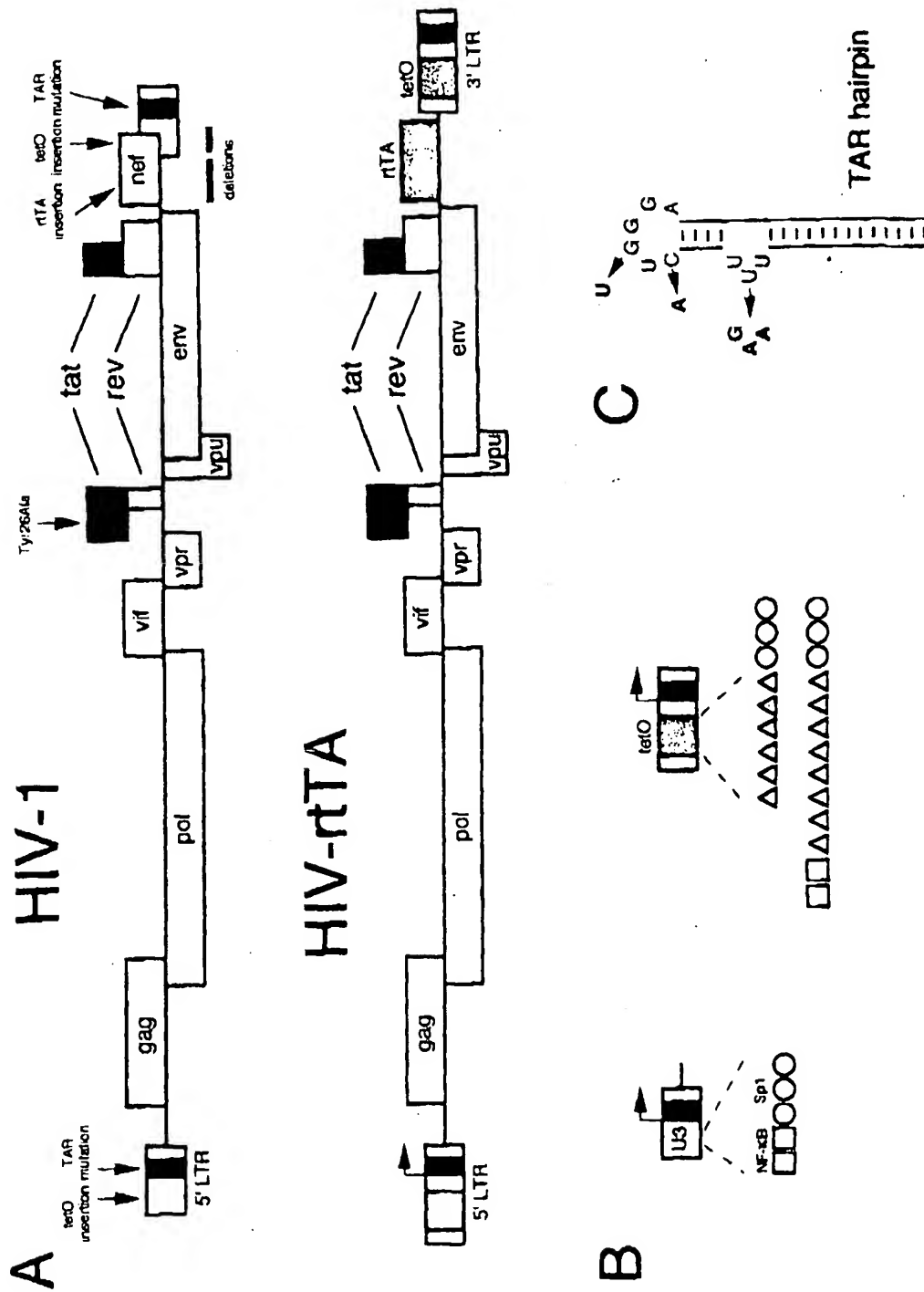


Fig. 1

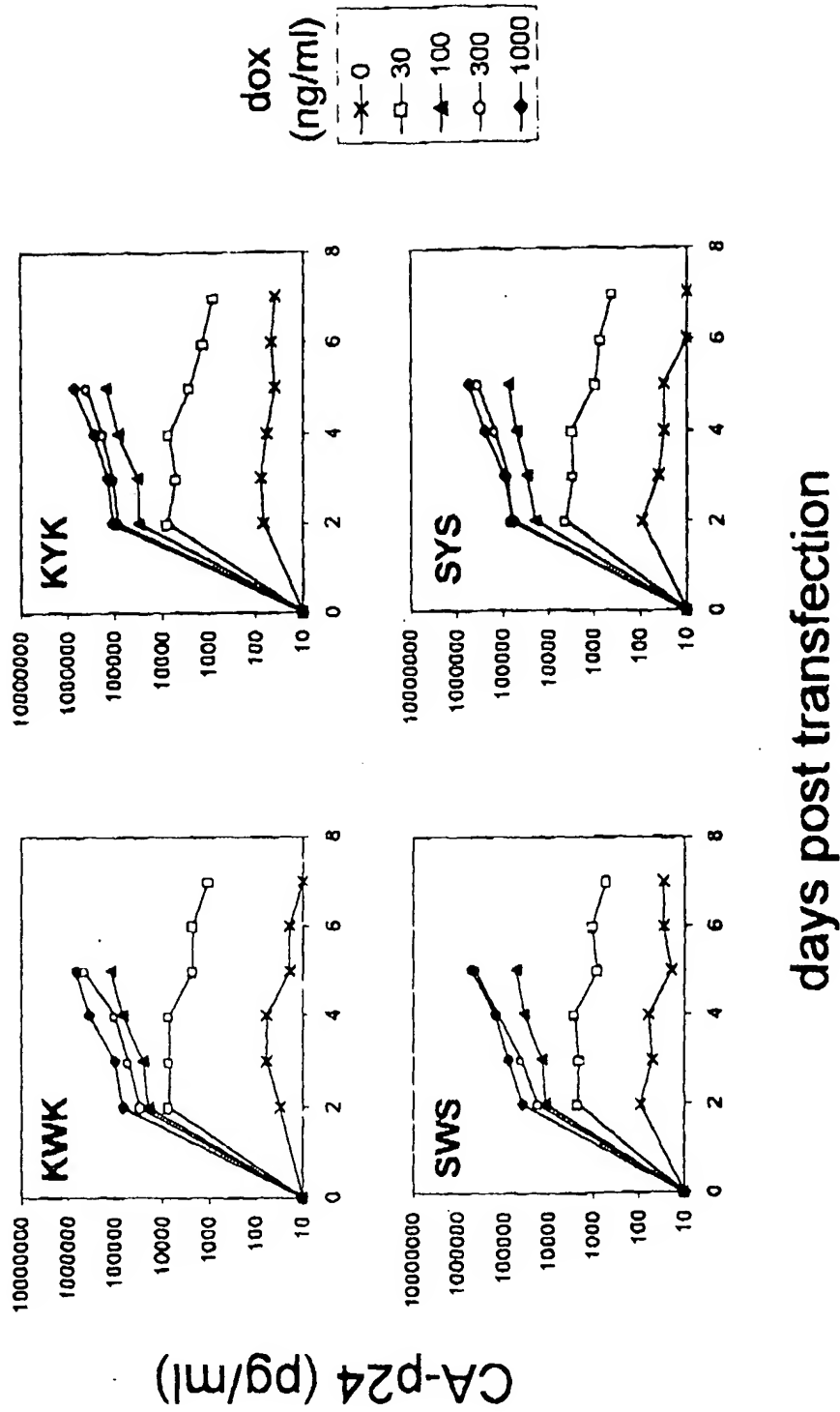


fig. 2

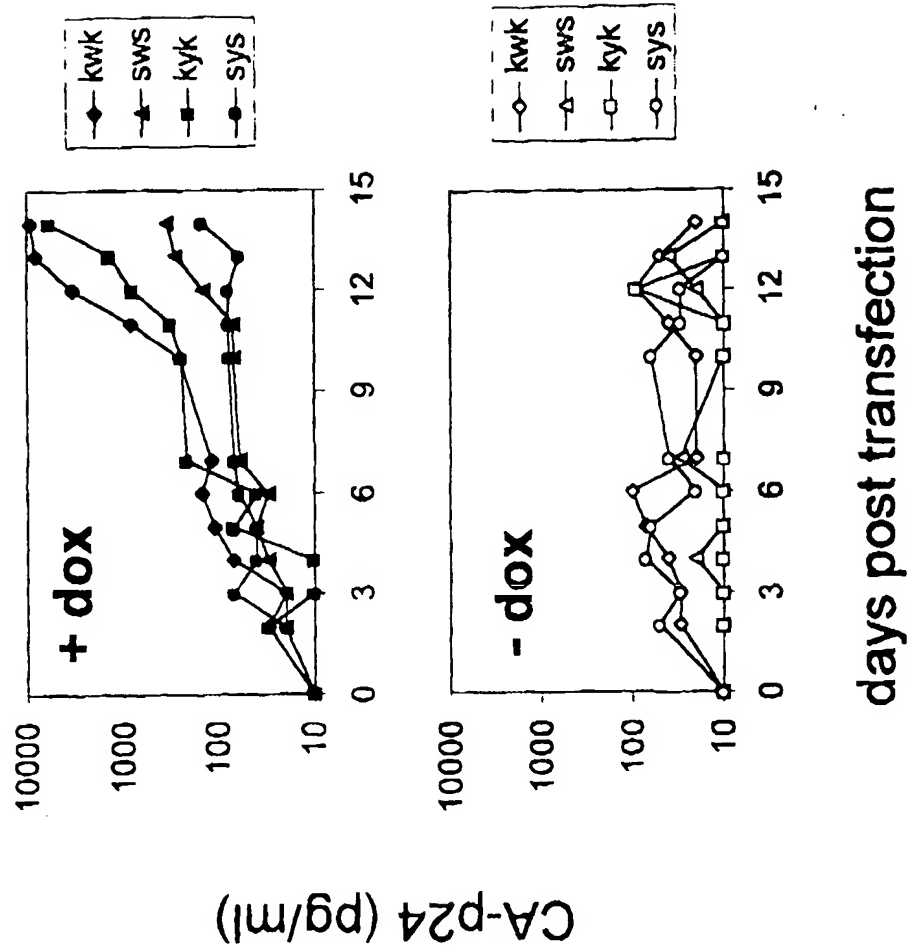


Fig. 3

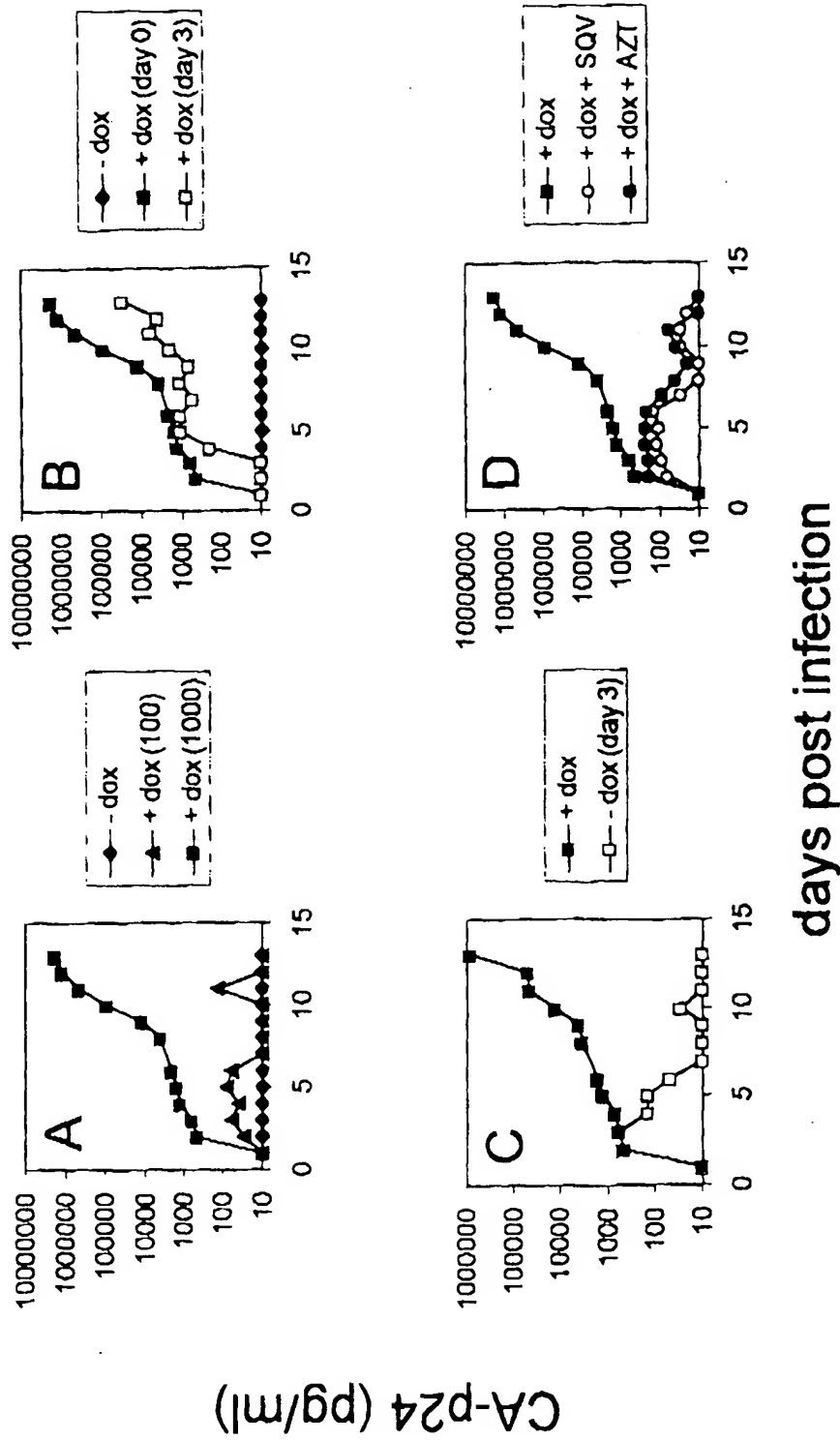


Fig. 4



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DOCUMENTS CONSIDERED TO BE RELEVANT			
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X	YAO F ET AL: "A novel tetracycline-inducible viral replication switch" HUM GENE THER, vol. 10, no. 3, 10 February 1999 (1999-02-10), pages 419-427, XP000877364 * figure 3B *	1,16,20, 26	
X	WO 97 48277 A (GAGE FRED H ;SUHR STEVEN T (US); SALK INST FOR BIOLOGICAL STUDI (U) 24 December 1997 (1997-12-24) * claims 4,13,21 *	1-5,7, 13,14, 16-22	
			TECHNICAL FIELDS SEARCHED (Int.Cl.7)
			C12N A61K
INCOMPLETE SEARCH			
<p>The Search Division considers that the present application, or one or more of its claims, does/do not comply with the EPC to such an extent that a meaningful search into the state of the art cannot be carried out, or can only be carried out partially, for these claims.</p> <p>Claims searched completely :</p> <p>Claims searched incompletely :</p> <p>Claims not searched :</p> <p>Reason for the limitation of the search:</p> <p>Although claim 27 is directed to a method of treatment of the human/animal body (Article 52(4) EPC), the search has been carried out and based on the alleged effects of the compound/composition.</p>			
Place of search		Date of completion of the search	Examiner
THE HAGUE		7 March 2000	Lonnoy, O
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